## **Temporal stability of therapeutic microbubbles**

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## **Background, Motivation and Objective**

Non-invasive blood-brain barrier (BBB) opening using focused ultrasound (FUS) requires intravenous injection of pre-formed microbubbles. Although microbubble behavior during exposure to imaging sequences has been studied extensively, microbubble stability within a therapeutic field remains relatively unexplored. Here, we studied the temporal stability of microbubbles during therapeutic FUS exposure over two timescales: the short time scale (i.e.,  $\mu$ s of low-frequency ultrasound exposure) and the long time scale (i.e., days post-activation). Our objective was to test whether in-house lipid-shelled microbubbles maintained their capacity to produce similar BBB opening for a period of up to 3 weeks following activation.

## Statement of Contribution/Methods

In-house manufactured lipid-shelled microbubbles (DSPC:DSPE-PEG2000 molar ratio 9:1,  $C_4F_{10}$  gas core) were first characterized in terms of their size and concentration using optical microscopy. They were then channeled to flow through a 4-mm vessel within a tissue-mimicking phantom (5% gelatin) and were exposed to therapeutic pulses (fc: 0.5 MHz, peak-negative pressure: 300 kPa, pulse length: 1 ms, pulse repetition frequency: 1 Hz, n=10). We recorded and analyzed the microbubble acoustic emissions with concentration-matched samples (10<sup>7</sup> microbubbles/ml) on day 0, 7, 14, and 21 after activation. *In vivo* experiments were conducted in mice (n=3) using the same parameters and at the same time-points in order to examine the therapeutic efficacy of microbubbles in BBB opening over time.

## **Results/Discussion**

Microbubbles had a concentration decay constant of  $0.02 \text{ d}^{-1}$  but maintained a stable size distribution for up to 3 weeks (< 10% variation). Temporal stability decreased while inertial cavitation increased over time both in vitro and in vivo, possibly due to changes in the lipid shell. BBB opening volume in mice (n=3) measured through T<sub>1</sub>-weighted contrast-enhanced MRI was equal to  $19.1 \pm 7.1 \text{ mm}^3$ ,  $21.8 \pm 14 \text{ mm}^3$ ,  $29.3 \pm 2.5 \text{ mm}^3$ , and  $38 \pm 20.1 \text{ mm}^3$  on day 0, 7, 14, and 21, respectively, showing no significant difference over the long time scale (p-value: 0.49). In conclusion, microbubbles maintain their capacity to produce similar therapeutic effects over a period of 3 weeks after activation, as long as the natural concentration decay is accounted for.



Figure: A) In vitro and in vivo experimental setup. B) In vitro microbubble stability. i) Mean and maximum microbubble size evolution. ii) Microbubble lifetime over time, expressed as the time required for 20% or 80% of the total energy to be emitted (i.e., t20 and t80). iii) Normalized spectra over time. iv) Cavitation doses evolution. SCDh: harmonic stable cavitation dose, SCDu: ultraharmonic cavitation dose, ICD: inertial cavitation dose. C) In vivo microbubble stability. Spectrogram evolution, showing increase of broadband emissions with storace time. D) BBB opening evolution over time.